

# EXPERT OPINION

1. Introduction
2. Drugs used for topical treatment of CL
3. Skin structure and skin permeability in CL lesions
4. Conventional drug delivery systems
5. Novel drug delivery systems
6. Conclusion
7. Expert opinion

## Drug delivery systems for the topical treatment of cutaneous leishmaniasis

Guilherme Carneiro, Marta Gontijo Aguiar, Ana Paula Fernandes & Lucas Antônio Miranda Ferreira<sup>†</sup>

<sup>†</sup>Federal University of Minas Gerais, Department of Pharmaceutics, Faculty of Pharmacy, Belo Horizonte, Minas Gerais, Brazil

**Introduction:** The parenteral administration of pentavalent antimonials for the treatment of all forms of leishmaniasis, including cutaneous leishmaniasis (CL), has several limitations. Therapy is long, requiring repeated doses and the adverse reactions are frequent. Topical treatment is an attractive alternative for CL, offering significant advantages over systemic therapy: fewer adverse effects, ease of administration, and lower costs.

**Areas covered:** This review covers, from 1984 to the present, the progress achieved for the development of topical treatment for CL, using different drugs such as paromomycin (PA), imiquimod, amphotericin B (AmB), miltefosine, and buparvaquone. PA is the most commonly studied drug, followed by AmB and Imiquimod. These drugs were incorporated in conventional dosage forms or loaded in lipid nanocarriers, which have been used mainly for improved skin delivery and antileishmanial activity.

**Expert opinion:** Developing an effective topical treatment for CL using these antileishmanial drugs still remains a great challenge. Insights into the most promising delivery strategies to improve treatment of CL with PA and AmB using conventional dosage forms, lipid nanocarriers, and combined therapy are presented and discussed. The results obtained with combined therapy and alternative delivery systems are promising perspectives for improving topical treatment of CL.

**Keywords:** amphotericin B, cutaneous leishmaniasis, lipid nanocarriers, liposomes, paromomycin, skin penetration, topical treatment

*Expert Opin. Drug Deliv.* (2012) 9(9):1083-1097

### 1. Introduction

Leishmaniasis is caused by protozoan parasites that are transmitted by the bite of phlebotomine sandflies that injects infective metacyclic promastigotes under the vertebrate host skin. In vertebrate hosts, *Leishmania* survive and multiply as non-motile amastigotes in cells of the phagocytic mononuclear system at the site of infection [1]. Eventually, depending on several factors, parasites may spread to other cutaneous sites or to visceral organs [2]. Therefore, upon infection, patients may develop symptoms that range in severity from skin lesions to serious disfigurement and fatal systemic infection [3]. Based on this, leishmaniasis has been classified into two major clinical entities: i) tegumentary, which comprises cutaneous, cutaneous diffuse, and mucocutaneous leishmaniasis (MCL) and ii) visceral leishmaniasis (VL). Each syndrome depends on both the infecting parasite species and the host immune responses [3-5].

Cutaneous leishmaniasis (CL) results in formation of skin ulcers at the site of the sandfly bite. CL is caused mainly by *Leishmania (Leishmania) major*, *L. (L.) tropica*, *L. (L.) aethiopica* in the Old World, and by *L. (Viannia) braziliensis*, *L. (V.)*

**informa**  
healthcare

**Article highlights.**

- General information is provided about cutaneous leishmaniasis, its causative organisms, its symptoms, and the methods of conventional (pentavalent antimonials) or alternative (topical) treatment of this disease.
- Key information is arranged about the chemistry of the main drugs used for the topical treatment of cutaneous leishmaniasis, the actual knowledge of their mechanism of action, and their pharmacokinetics parameters.
- The structure and permeability of the healthy skin as well as changes in skin permeability during the progression of the disease and skin conditions in the topical treatment of the cutaneous leishmaniasis are discussed.
- The main conclusions of clinical studies or studies on animals using conventional dosage forms containing topical drugs used for the treatment of cutaneous leishmaniasis are presented.
- The advantages of using lipid nanocarriers loaded with paromomycin and amphotericin B for the topical treatment of cutaneous leishmaniasis are presented.

This box summarizes key points contained in the article.

*guyanensis* L. (*V.*) *panamensis*, *L. (L.) amazonensis* and *L. (L.) mexicana* in the New World. The disease is usually self-limiting, but infection by New World species, for example, may spread to lymph nodes and other cutaneous sites. It is generally assumed that MCL, often caused by *L. (V.) braziliensis*, develops after cure of the initial skin lesions. The late development of metastatic lesions leads to partial or total destruction of the mucous membranes, progressing to severe deformities or even death [3].

Considering the diversity of clinical manifestations, immunopathological responses, and parasite species associated with leishmaniasis, the concept that leishmaniasis may be more properly defined as a diverse group of syndromes has recently emerged [3] and, certainly, has implications for development of effective treatment alternatives.

Meglumine antimoniate and sodium stibogluconate have been used for more than half of a century in the therapy of all forms of leishmaniasis. These drugs have to be given parenterally, daily, for at least 3 weeks (typically, 20 mg/kg/day for 20 – 30 days). The treatment with antimonials is long, requiring repeated doses. The adverse reactions (arthralgia, myalgia, nausea, vomiting, elevated liver and pancreatic enzymes, etc.) are frequent, the most serious one on the cardiovascular system [6,7].

Alternative therapies for the treatment of all forms of leishmaniasis have been intensively investigated, including the identification of drugs and formulations that can be administered topically for the treatment of CL [8,9].

Topical treatment is an attractive alternative for CL [6]. Topical formulations offer significant advantages over systemic therapy: fewer adverse effects, ease of administration and reduced cost. Among the different drugs that have been

studied for the topical treatment of CL are paromomycin (PA), imiquimod, amphotericin B (AmB), miltefosine, and buparvaquone. PA is the most commonly studied drug for topical treatment of CL caused by different species [10-12].

Most studies evaluating formulations containing antileishmanial drugs for topical treatment of CL have investigated conventional dosage forms such as ointments, creams, and gels. Although experimentally infected animal studies have shown promising results, clinical trials have shown variable efficacy and sometimes disappointing results [13,14]. This may be attributed, at least in part, to low skin penetration of antileishmanial drugs. Therefore, novel drug delivery systems such as lipid nanocarriers present the potential to improve drug penetration into or across skin [15,16].

Among the lipid nanocarriers of antileishmanial drugs, liposomes, the most extensively investigated, provide improved efficacy, probably due to enhanced drug skin penetration [16]. When applied to the intact skin, liposomes can enhance dermal or transdermal drug penetration depending on the composition and size of the vesicles [17]. Different mechanisms of action for liposomes as skin drug delivery systems have been suggested: penetration enhanced by liposome components individually; vesicle adsorption to and/or fusion with the stratum corneum (SC); intact vesicle penetration into and through the intact skin; and follicular penetration (Figure 1) [18].

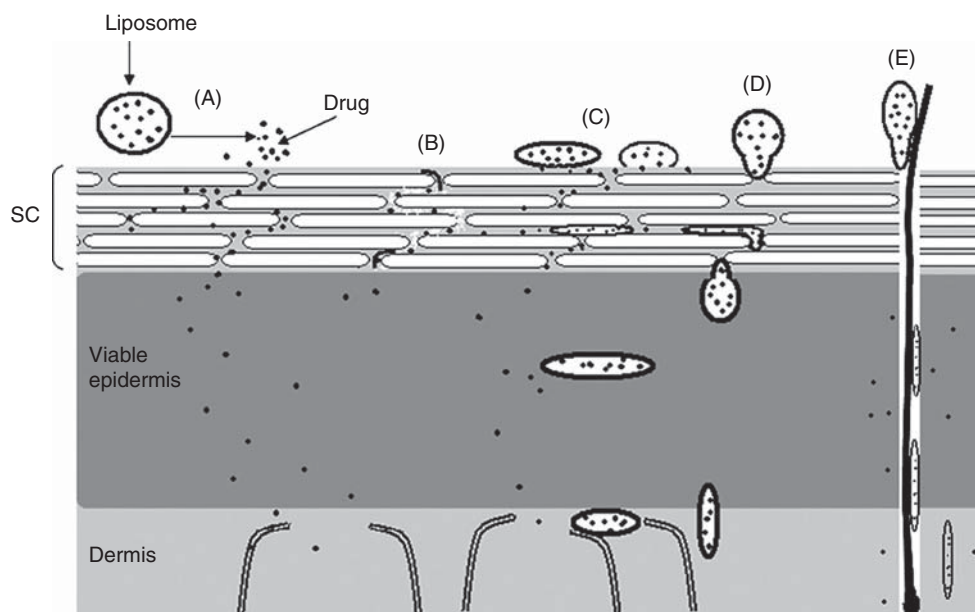
The rationale for the use of liposomes in the topical treatment of CL when compared with conventional formulations can be schematized as shown in Figure 2. When applied to the skin with normal or impaired barrier, liposomes can increase the drug skin penetration [16-19]. In addition, when applied to the totally damaged skin (open lesions or full-thickness skin wound), liposomes showed the ability to accelerate wound healing when compared to conventional formulations [20-22].

Following a brief description of the more studied drugs for the topical treatment of CL, as well as on the nature of the CL lesion and intact skin barrier, this review critically evaluates conventional dosage forms and novel drug delivery systems, concentrating primarily on lipid nanocarriers used for the topical treatment of CL.

## 2. Drugs used for topical treatment of CL

### 2.1 Paromomycin (PA)

The aminoglycoside antibiotic PA is a highly hydrophilic drug with a high molecular weight and relative lipid insolubility (Table 1). PA has variable activity against different *Leishmania* species [23]. Studies on PA have demonstrated that it was able to inhibit protein synthesis *in vivo*, though in the promastigote stage of the parasite [24]. PA has low bioavailability when administered orally. When administered intramuscularly, PA is quickly absorbed, reaching peak plasma levels within 1 h [25,26]. The nephrotoxicity and ototoxicity, adverse events commonly associated with the



**Figure 1. Possible mechanisms of action of liposomes as skin drug delivery systems.** (A) is the free drug mechanism, (B) is the penetration enhancing process of liposome components, (C) indicates vesicle adsorption to and/or fusion with the stratum corneum (SC) and (D) illustrates intact vesicle penetration into or into and through the intact skin (not to scale).

Reproduced with permission of Elsevier from [18].

aminoglycosides, occurred in about 1.4% of patients, after parenteral administration [23]. The PA percutaneous absorption after topical application was investigated in ulcerated CL lesions of BALB/c infected mice. The PA concentration in plasma after 1 h ranged from 2 to 22  $\mu\text{g/mL}$ , suggesting that topically applied PA can be absorbed systemically [27].

## 2.2 Miltefosine

Miltefosine, an alkylphosphocholine (Table 1), originally developed for the topical treatment of cutaneous metastases of breast cancer, was the first oral drug with antileishmanial activity. Miltefosine is registered for the oral treatment of VL in India and Germany, and for CL in Colombia [28,29]. Miltefosine can act through different mechanisms on *Leishmania* spp., altering the metabolism of lipids and inducing the production of interferon  $\gamma$  (IFN- $\gamma$ ) [30,31]. Adverse reactions of miltefosine over the gastrointestinal system include nausea, vomiting, and diarrhea [32-34]. Toxicity studies have shown that the other organs most affected are the eyes, kidneys, and the reproductive system [7].

## 2.3 Amphotericin B

AmB is a highly hydrophobic drug with a high molecular weight that exhibits amphiphilic and amphoteric properties (Table 1). It is a polyene antifungal produced by *Streptomyces nodosus* that was introduced originally as a drug to combat systemic fungal infections. AmB also exhibits antiparasitic activity and is a second-line therapy useful for the treatment of CL, VL, and MCL [35-37]. The possible mechanism of

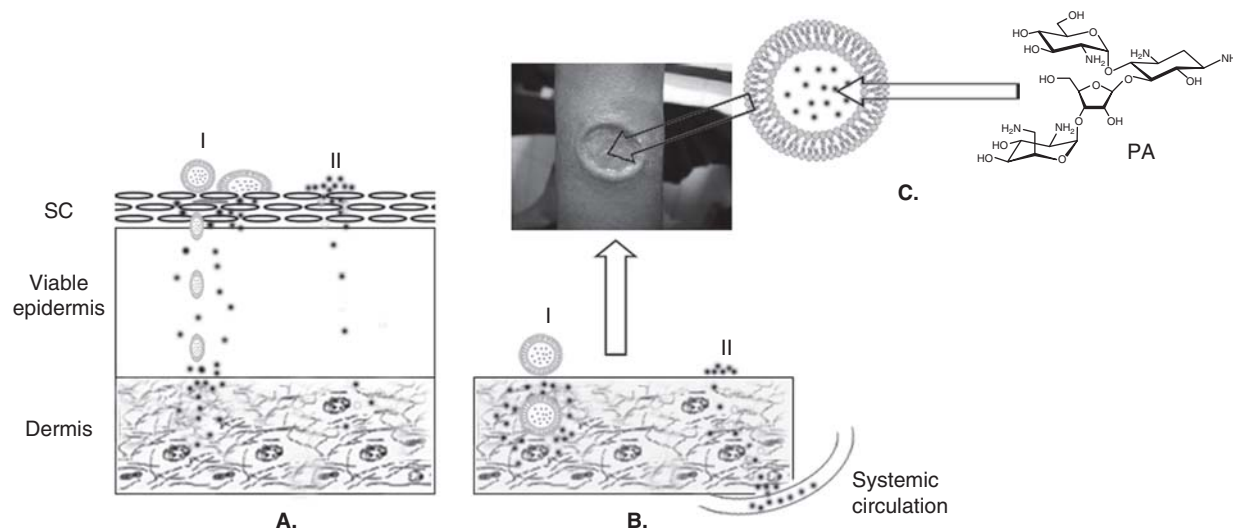
action of AmB over *Leishmania* occurs through the formation of channels that are permeable to small cations and anions [36,38]. However, medical application of AmB is limited by its toxicity. Required hospitalization, constant monitoring of patients, prolonged duration of treatment, and infusion-related adverse events are the drawbacks of AmB deoxycholate [39].

## 2.4 Imiquimod

Imiquimod, a nucleoside analogue of the imidazoquinoline family, was the first immune response modifier used for the treatment of infectious skin conditions (Table 1). It was approved originally for the topical treatment of external anogenital warts [40]. There are case reports and preliminary studies suggesting its effectiveness in the treatment of CL [41,42]. The effects of imiquimod on innate immune responses, via toll-like receptors, suggest a potential antileishmanial activity that was demonstrated by inducing the release of nitric oxide [43,44]. Less than 1% of the drug is recovered in urine after topical application. Topical imiquimod (5% cream) is only mildly irritating and does not lead to systemic toxic effects [43].

## 2.5 Buparvaquone

Buparvaquone (BPQ), a hydroxynaphthoquinone (Table 1), is currently marketed for the treatment of theileriosis in cattle (as Butalex<sup>®</sup>). It has several physicochemical properties suitable for topical delivery (low molecular weight, low melting point, etc.). However, it has low aqueous solubility and high



**Figure 2. Rationale for the use of liposomes in topical treatment of CL.** Liposomes (I) or free drug (II) can be applied either to the intact skin (A) or to the ulcerated skin (B). When applied on the intact skin, liposomes can improve drug penetration in comparison with the free drug; when applied on the highly damaged skin, liposomes provide targeting skin avoiding systemic absorption. Hydrophilic drugs such as PA loaded in liposomes can be applied in the leishmaniasis lesions (C), in which skin is ulcerated.

PA: Paromomycin; SC: Stratum corneum.

lipophilicity. Attempting to increase aqueous solubility and absorption in topical treatment for CL, a phosphate prodrug approach has been investigated [45,46].

### 3. Skin structure and skin permeability in CL lesions

Human skin is composed of two parts: the epidermis and the dermis. The superficial layer, and also the thinner one, is the epidermis, whereas the deeper layer is the dermis. The epidermis or, specifically, the SC, its outermost layer, is considered the main barrier to the passage of substances through the skin.

The SC is formed by dense, functionally dead, keratinized cells, named the corneocytes. These cells are surrounded by the intercellular lipid matrix composed of non-polar lipids organized in lamellar lipid layers. Thus, the SC is mainly a hydrophobic layer. Although the SC is 10 – 20  $\mu\text{m}$  thick, it is also considered responsible for the barrier function of the skin. It hampers the skin penetration of drugs, other chemicals, and microorganisms besides being involved in the regulation of transepidermal water loss [47,48].

Three possible pathways for drug skin delivery have been considered: the intercellular route, between the corneocytes, sinuously through the lipid matrix; the transcellular route, through the corneocytes and the lipid matrix; and the transappendageal route, through the cutaneous appendages (hair follicles, sebaceous and sweat glands). The intercellular route is considered the most important one to drug penetration, since on the transcellular route, the drug has to cross the skin passing through both the lipid structures and the

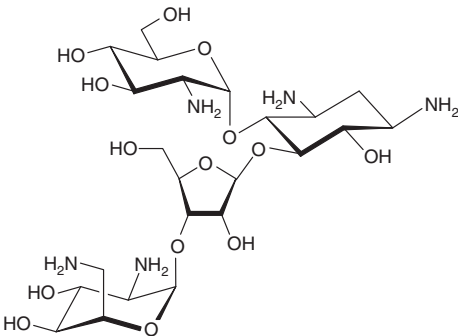
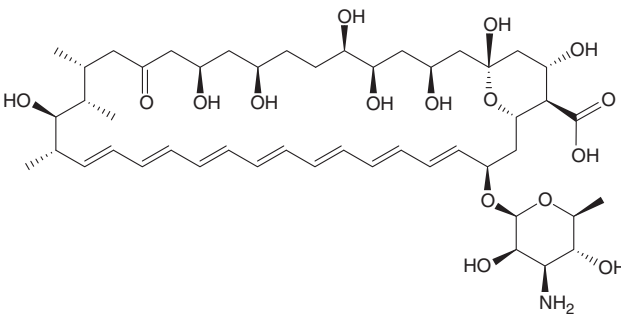
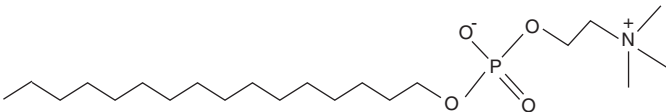
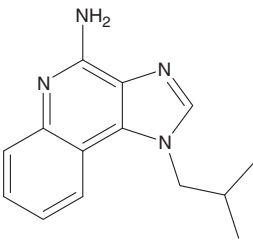
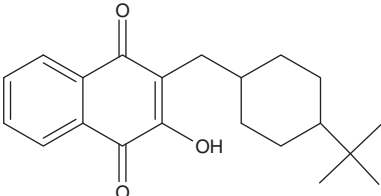
cytoplasm of the corneocytes [49,50]. Although cutaneous appendages occupy less than 0.1% of the total skin surface, this path can circumvent the low diffusivity of the SC, acting as a diffusional shunt [47,49].

To facilitate drug absorption through the skin, it is also important to select an adequate molecule. An ideal molecule would have to possess a low molecular mass, preferably less than 600 Da; solubility in oil and water, i.e., balanced partition coefficient; and a low melting point [15]. Thus, the skin penetration of drugs such as PA, which is highly hydrophilic and has a high molecular weight, is hampered.

In CL, the permeability of the skin is highly variable in comparison to the normal skin. CL typically begins with a small erythema and swelling which increases in size at the area where an infected sandfly has bitten the host. Then, the erythema develops into a papule, which increases in size, becoming a nodule that progressively ulcerates over a variable period ranging from 2 weeks to 6 months. After this time, it becomes the characteristic lesion of CL [51,52], with the loss of epidermis and part of the dermis. Therefore, since the skin is damaged, the barrier function exerted by the SC is absent and the absorption of any kind of drug used in topical treatment can be enhanced, especially the hydrophilic ones [53].

The size and evolution of lesions vary with the infecting strain of *Leishmania*. Old World CL and *L. mexicana* lesions tend to heal spontaneously after 6 – 12 months, but *L. braziliensis* may take years to heal. New World CL tends to be more severe and spontaneous healing is rare, with lesions which vary widely, ranging from small, dry lesions to large ulcers. In severe disease, keratotic nodules can also develop. Both in

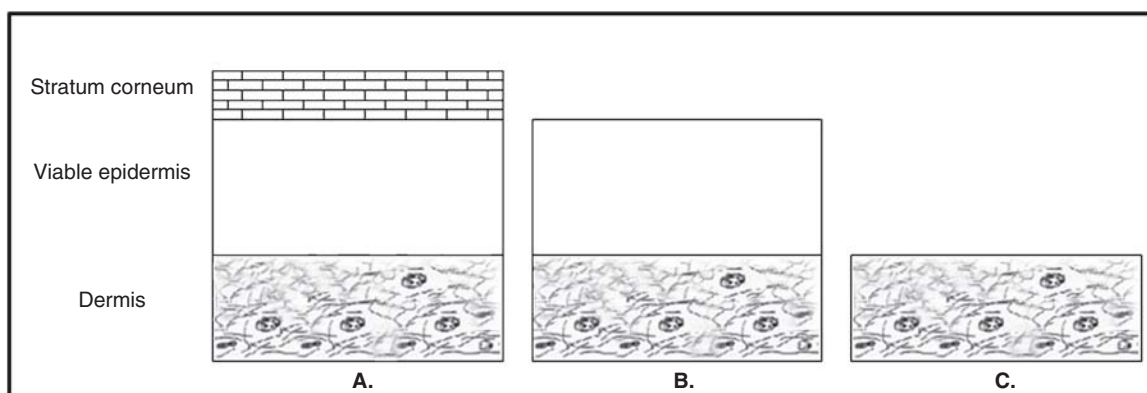
**Table 1. Structural formula and physicochemical characteristics of drugs used for the topical treatment of CL.**

Drug	Molecular structure	Physicochemical characteristics
Paromomycin		Very soluble in water, insoluble in alcohol, chloroform and ether [99]
Amphotericin B		Insoluble in water, soluble dimethyl sulfoxide and propylene glycol, very slightly soluble in methanol [100]
Miltefosine		Solubility: > 10 g/100 mL in water, methanol or ethanol; < 10 g/100 mL in: acetone, n-Pentane [33]
Imiquimod		Crystalline odorless solid, poorly soluble in water [43]
Buparvaquone		Solubility in water: < 0.03 µg/mL, in pH 3.0, 5.0 or 7.4 [88]

keratotic nodules and in scar tissues formed during the treatment, the SC barrier function is recovered or even more pronounced. Thus, the drug skin penetration may become hindered at the end of treatment in healing lesions [52,54].

Therefore, skin conditions represent important considerations in the topical treatment of CL. Formulations can be applied either to thickened lesions, with an additional barrier

to absorption or, more commonly, to open lesions (e.g., ulcers), in which the barrier properties of the epidermis have been completely lost [6]. In addition, the topical treatment of CL can last for 20 – 30 days [8]. Therefore, re-epithelization of open lesions can be observed, at least partially, during therapy, whereas the renewal of the epidermis contributes to a decrease in drug penetration due



**Figure 3. Schematic diagram of the skin conditions during the CL.** Formulations can be applied to thickened lesions, in which the barrier properties are normal or even higher than the intact skin, or to open lesions, in which the barrier properties of the epidermis have been completely lost. Application on partially damaged skin (skin without SC) is also possible. (A) Intact skin: with SC, viable epidermis and dermis; (B) Partially damaged skin: without SC; (C) Highly damaged skin: tissue without epidermis (open leishmaniasis lesion; ulcerated skin).

SC: Stratum corneum.

to barrier repair. An ideal formulation for the topical treatment of CL should be used to provide improved topical delivery in all antagonistic situations (i.e., intact, partially, or completely damaged barrier) (Figure 3). Finally, drug delivery systems should promote drug penetration into the deeper dermal layer, as the parasites in CL are mainly located in the dermis.

#### 4. Conventional drug delivery systems

Ointments, creams, and hydrophilic gels are the conventional drug delivery systems most frequently tested for the topical treatment of CL.

##### 4.1 Conventional PA delivery systems

Petrolatum ointments, containing PA associated with methylbenzethonium chloride (MBCL), an ammonium quaternary surfactant, which also has antileishmanial activity, have been evaluated with favorable results, in a series of studies in animal models infected with different species causing CL [55-58]. Several clinical trials have also been performed to test this formulation in CL patients from different geographical regions of the Old and New World (Table 2) [59-62].

However, MBCL induces local irritation, which has prevented its use in some cases [55,57]. The local side effects due to MBCL have led to the development of alternative topical formulations, replacing the MBCL by urea, which has lower toxicity when compared to MBCL. The ointment PA/urea cured all *L. major* lesions on BALB/C mice [13]. However, clinical trials have shown that this formulation is not as effective as that containing PA/MBCL (Table 2) [10,14,63-65].

Replacement of MBCL by addition of 0.5% of gentamicin has also been evaluated as an alternative PA topical formulation (cream) [58]. Based on the data obtained in

animal models, studies were conducted to evaluate the clinical efficacy [8,12]. However, variable cure rates were reported in comparison to placebo (Table 2) [8,12].

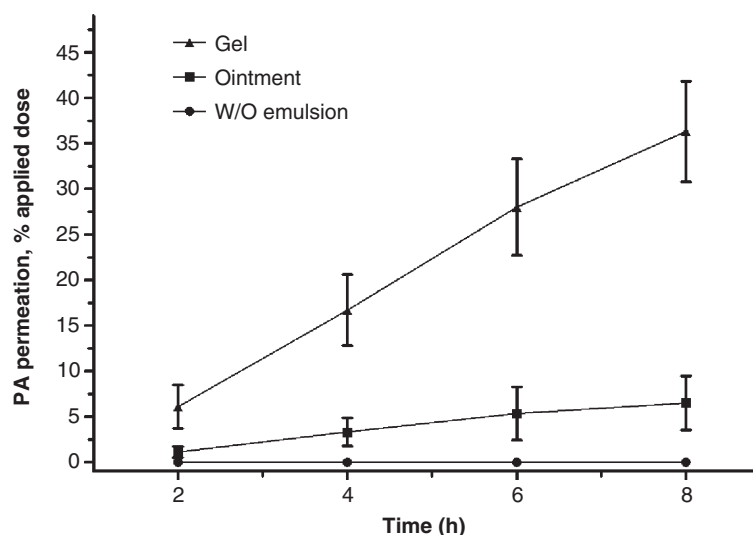
Considering that parasites in CL are located mainly in the dermis, drug penetration into and across skin would be desirable. To investigate the influence of different formulations (oil/water cream, ointment and hydrophilic gel) on the skin permeation of PA, *in vitro* studies on Franz cells were performed. As an attempt to mimic the conditions of skin lesions observed in CL, the permeation studies were conducted using two skin models: intact mouse skin and stripped mouse skin (removal of the SC), which was used to mimic the damaged skin. PA permeation across intact skin was low, regardless of the formulation tested. When the SC was removed, PA permeation from hydrophilic formulations (cream and gel) was enhanced significantly [66]. However, studies were carried out on hairless mice skin, a model that cannot be easily extrapolated to human skin [67]. Thus, additional permeation studies were performed using pig ear skin, a more relevant model for human skin [68]. Three models were tested: intact and stripped skin and dermal membranes (pig skin without epidermis). The dermal membrane has been considered a good model for ulcerated skin [69], which is frequently observed in CL. PA permeation across intact pig skin was low, regardless of formulation tested (gel, water/oil emulsion and ointment) (data not published). In contrast, PA permeation across dermal membranes from hydrophilic gel was higher than that observed for other formulations (Figure 4). These findings are consistent with data obtained previously, which showed that PA penetration into or across skin from gels was higher than that observed for hydrophobic formulations.

Since CL lesions often develop into an ulcer without epidermis, one may consider that the principal barrier to

**Table 2. Animal studies and clinical trials using conventional dosage forms containing PA for topical treatment of CL.**

Concentration (%)	Formulation	Composition	Studies in animals	Clinical trials
15	Hydrophobic ointment	White soft paraffin + 12% MBCL	[42,55-58]	[10,59-62]
15	Hydrophobic ointment	White soft paraffin + 10% urea	[13]	[10,14,63-65]
15	Cream (WR 279.296)	0.5% gentamicin in a cream vehicle	[58]	[8,12]
10	Hydrogel	Hydroxyethylcellulose	[27,72-74]	[11]

CL: Cutaneous leishmaniasis; PA: Paromomycin; US: United States.



**Figure 4. *In vitro* penetration of PA from topical formulations (gel, ointment and w/o emulsion) containing PA at 1% across dermal membranes.** The formulations were applied over biopsies of pig skin without epidermis (dermal membranes) mounted in Franz cells. Each point represents means  $\pm$  SD (n = 3).

PA: Paromomycin; SD: Standard deviation; W/O emulsion: Water-in-oil emulsion.

drug skin penetration is absent [70]. For the hydrophilic gel, PA is dissolved and available allowing its prompt release and penetration into or across the skin. In contrast, PA is insoluble in hydrophobic ointments and therefore drug release depends strongly on the particle size. For the water/oil emulsion, although the drug is soluble in the internal aqueous phase, drug diffusion across the external oil phase is the rate-limiting step. Lalor *et al.* have shown that, in an emulsion, the interaction of the drug between the aqueous and oily phase can determine the extent in which the thermodynamic activity is decreased in the external phase in contact with the membrane [71].

These findings prompted investigations on the efficacy of this new formulation (hydrophilic gel) in animal models. Accordingly, the studies performed have demonstrated that a PA gel formulation was highly effective to treat lesions in *L. (L.) major*, *L. (L.) amazonensis*-infected mice or *L. (V.) braziliensis*-infected hamsters [27,72,73]. This formulation was also recently tested in CL patients infected by *L. (V.) braziliensis* that could not be submitted primarily to meglumine antimoniate therapy [11]. Treatment was well tolerated and

patients who did not present healing after a 20-day course continued spontaneously treating themselves. The average time of treatment was 30 days and a cure rate of 60% (9/15) was observed. These findings suggest that the efficacy of this formulation should be further evaluated in clinical trials.

Efficacy of the topical treatment of CL may not be attributed, however, solely to drug skin delivery. Other variables may significantly affect cure rates and reduction of tissue parasitism. *Leishmania* species differ significantly in their susceptibility to distinct drugs, including PA. In addition, based on the fact that infection by New World *Leishmania* species may disseminate reaching other distant body sites, combined alternative therapies have been advocated [37]. In agreement, findings from pre-clinical trials revealed that combined topical PA-gel and oral miltefosine is highly effective regardless of the parasite species susceptibility to both drugs, promoting complete elimination of parasites in infected mice at the site of infection as well as in internal organs [27,74].

Finally, although the drug penetration has been improved through the use of the gel, it is noteworthy that PA

permeation across the stripped pig skin, a damaged skin model, was still quite low. In this context, drug nanocarriers such as liposomes have demonstrated the capacity to increase drug penetration into and across the skin, when compared to conventional formulations [75,76]. In addition, application on partially damaged tissue (stripped skin) provided targeted, sustained topical delivery, [77] whereas on open lesions, liposomes, as compared to conventional formulations, showed the ability to accelerate wound healing [20-22]. Thus, PA-loaded liposomes could provide a reliable alternative for the topical treatment of the CL.

#### 4.2 Conventional AmB delivery systems

A topical preparation of Fungizone® (conventional formulation of AmB) in white paraffin associated with the MBCL was evaluated in experimental CL. However, this formulation was not effective [55]. This could be explained by the fact that to obtain effective skin penetration of AmB, an adequate drug-carrier system is required. Thus, a drug-carrier system loaded with AmB may also prove to be an effective alternative for the topical treatment of CL.

#### 4.2 Conventional miltefosine delivery systems

The potential of oral miltefosine against CL has been demonstrated [78,79]. In agreement, Miltex®, a micellar solution containing miltefosine at 6%, which was originally used for the treatment of cutaneous metastases of breast cancer, efficiently reduced the parasite burden in experimentally infected mice after topical administration [80].

#### 4.3 Conventional imiquimod delivery systems

The imiquimod at 5%, which is available as a cream (Aldara®), has demonstrated antileishmanial activity for the treatment of CL, both in experimentally infected mice [42,81] and in humans [41,82-85]. These studies suggest that Imiquimod in combination with pentavalent antimonials presents potential to reduce the therapy course with antimony alone. However, the results of human clinical trials of Aldara®, as a primary treatment for CL, were of limited success [43,86].

#### 4.4 Conventional buparvaquone delivery systems

Different formulations (solution, gels, and w/o emulsion) containing BPQ and 3-phosphonooxymethyl-buparvaquone (3-POM-BPQ), a prodrug of BPQ, were evaluated in the topical treatment of CL. The prodrug was rapidly hydrolyzed in the skin, being used to improve aqueous solubility and enhance skin absorption [87]. Studies of *in vitro* permeation demonstrated that BPQ and 3-POM-BPQ can penetrate the skin when applied in different formulations. The BPQ penetration was highest from isopropyl myristate, hydrophilic gel, and w/o emulsion, whereas 3-POM-BPQ penetration was greatest from an anhydrous gel [88]. The efficacy of these formulations in *in vivo* models of CL was evaluated [45]. The formulations showed antileishmanial activity reducing the cutaneous parasite burden and lesion size.

### 5. Novel drug delivery systems

The topical treatment of CL using novel drug delivery systems consists of the incorporation of the drug in nanostructured carrier systems (NCS), i.e., complex particles in the nanometer range. NCS have been proposed with the main objective to enhance skin penetration, to protect the drug against degradation, and to promote controlled release and follicular targeting. Lipid nanocarriers such as liposomes loaded with PA as well as nanoemulsions or lipid complexes (Amphocil®) loaded with AmB are the most frequently evaluated NCS for the topical treatment of CL.

Liposomes are spherical vesicles formed basically by phospholipids which associate themselves spontaneously and form a bilayer containing a centralized aqueous cavity (Figure 2). They are most commonly formed by phospholipids of natural, synthetic, or semi-synthetic origin and possess great potential for the delivery of hydrophilic, lipophilic, and amphiphilic drugs. The hydrophilic drugs can be encapsulated in the aqueous cavity, while the lipophilic ones are located in the non-polar portion of the bilayers. Amphiphilic drugs can be loaded in the hydrophilic-lipophilic interface [89]. Depending on the composition of the phospholipids (saturated or unsaturated) and the presence of co-surfactants (cholesterol, sodium cholate), rigid, fluid or elastic vesicles can be obtained and dermal or transdermal delivery can be favored [15,90].

Nanoemulsions are lipid formulations constituted by oil and aqueous phases with droplet size in the nanometric scale. Surfactants and co-surfactants are often used to stabilize these systems. Due to their oil phase, nanoemulsions are well established carriers for lipophilic drugs [91]. Other nanocarriers studied include ribbon-like lipid complex with dimyristoylphosphatidylcholine and dimyristoylphosphatidylglycerol (Adelcet®) or even disc-like colloidal dispersions with cholesteryl sulfate (Amphocil®).

There are few studies involving lipid nanocarriers loaded with antileishmanial drugs for the topical treatment of CL. However, these systems have been proposed as carriers of hydrophilic PA or hydrophobic AmB, which are both high molecular weight drugs and, therefore, difficult to penetrate the skin.

#### 5.1 Liposomes loaded with PA

The first efforts to obtain PA encapsulation into liposomes were reported by Ferreira et al. [92]. The PA penetration from fluid liposomes such as large unilamellar vesicles (LUV), through intact and stripped mouse skin, was investigated. In intact skin, PA penetration into and across skin from LUV liposomes was significantly higher than that observed for the free drug solution. As expected, PA permeation across stripped skin was higher than that observed across intact skin, regardless of the tested formulation. Although permeation from solution was also high, liposomes promoted skin targeting when compared to the free drug. However, these studies have been

**Table 3.** PA permeation and penetration across pig skin from PC, PC/Chol liposomes and aqueous solution after 8 h.

Formulations	Intact skin		Stripped skin* (Permeation)	
	Penetration <sup>‡</sup>	Permeation	$\mu\text{g}/\text{cm}^2$	% applied dose
PC	$7.2 \pm 0.2$	ND	$14.6 \pm 0.6$	$19.6 \pm 0.8$
PC/Chol	$4.8 \pm 0.2$	ND	$4.5 \pm 0.4$	$6.1 \pm 0.5$
Solution	$1.9 \pm 0.1$	ND	$0.9 \pm 0.5$	$1.0 \pm 0.6$

Data represented as percent applied dose.

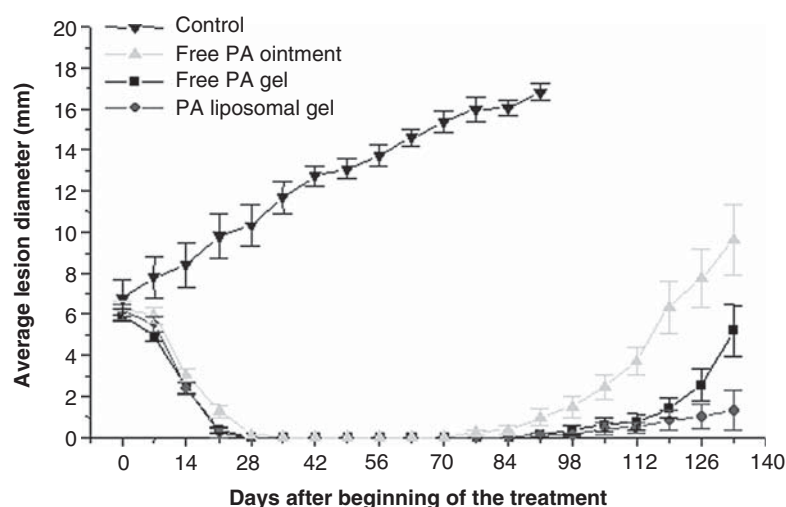
Reproduced with permission of Informa Healthcare from [19].

Values represent means  $\pm$  SD (n = 3 – 5).

\*SC was removed by stripping 30 times the skin with adhesive tape (Book Tape no. 845, Scotch 3M, EUA).

<sup>‡</sup>Penetration studies were not conducted with stripped skin.

Chol: Cholesterol; ND: Not detected; PC: Phosphatidylcholine; SC: Stratum corneum; SD: Standard deviation.



**Figure 5.** Evaluation of treatment efficacy in female BALB/c mice infected with *L. (L.) major*. Animals (n = 5) were treated topically with formulations containing PA at 5%: free PA ointment, free PA gel, PA liposomal gel or placebo gel for 12 days, twice a day. Lesion diameters are shown as averages and standard deviation. Mean lesion diameter was monitored for 133 days.

PA: Paromomycin.

carried out on hairless mice skin, a model that cannot be easily extrapolated to human skin [67]. Thus, more recently, the potential of liposomes in the delivery of PA was investigated by conducting permeation studies on pig ear skin, using two *in vitro* models: intact and stripped skin [19]. PA permeation across intact skin was low, regardless of the formulation tested. However, drug penetration into intact skin from liposomes was significantly higher than that from solution. Unexpectedly, PA permeation across stripped pig skin from solution was also low, but a remarkable increase in permeation was observed for PA-loaded liposomes (Table 3).

Topical treatment of *L. major* infected BALB/c mice resulted in a decrease in lesion size in animals treated with PA-loaded liposomes and free PA gel (Figure 5). However, local relapse, characterized by the reappearance of ulcers, occurred faster in animals treated with free PA than in those treated with liposomes. These findings suggest that liposomes

represent a promising alternative for the topical treatment of CL using PA [19].

Jaafari et al. also evaluated liposomes loaded with PA at 10% and 15%. Both liposomes promoted high permeation and retention of PA in the mouse skin [93]. These PA liposomal formulations were three to four times more effective against *L. major* amastigotes and promastigotes than PA solution. Treatment of *L. major* infected BALB/c mice with topical liposomes loaded with PA induced the complete cure of the lesions, and the mice had significantly reduced lower parasite burdens in the spleen than the control mice. However, in this study comparison of topical treatment with free PA was not reported.

## 5.2 Lipid nanocarriers loaded with Amphotericin B

Despite its amphiphilic nature, AmB has low solubility due to the asymmetric distribution of hydrophilic and hydrophobic groups and presents itself as a zwitterion compound [94].

**Table 4. *In vitro* permeation and penetration of AmB from different formulations across dermal membrane skin after 24 h.**

Formulation	Permeation	Penetration*	
		( $\mu\text{g}/\text{cm}^2$ )	(% applied dose)
Solution	< DL <sup>‡</sup>	17.5 $\pm$ 4	31 $\pm$ 5
Amphocil <sup>®</sup> ,§	< DL	15.2 $\pm$ 3	27 $\pm$ 4
Amphocil <sup>®</sup> ,¶	< DL	9.6 $\pm$ 3	17 $\pm$ 3
Fungizone <sup>®</sup>	< DL	3.5 $\pm$ 1	6 $\pm$ 2
FLmix	< DL	1.7 $\pm$ 0.3	3 $\pm$ 0.5
NE	< DL	1.1 $\pm$ 0.1	2 $\pm$ 0.15

Reprinted with permission from [98]. Copyright © American Scientific Publishers.

\*Values represent the mean  $\pm$  SD (n = 4).

<sup>‡</sup>DL = detection limit (0.02  $\mu\text{g}/\text{cm}^2$ ).

<sup>§</sup>Aqueous Amphocil<sup>®</sup>.

<sup>¶</sup>Hydroalcoholic Amphocil<sup>®</sup>.

These hydrophobic properties limit the skin penetration of AmB, mainly into the dermal layer. Lipid nanocarriers loaded with AmB constitute, therefore, a credible alternative to improve topical delivery.

Frankenburg et al. have reported the antileishmanial effects of topically applied lipid-based formulations loaded with AmB in *L. major* experimentally infected mice. The three tested preparations (Fungizone<sup>®</sup>, Amphocil<sup>®</sup>, and Adelcet<sup>®</sup>) were ineffective when applied topically, except when Amphocil<sup>®</sup> and Adelcet<sup>®</sup> were dispersed in 5% ethanol. No relapse was observed during the follow-up period after treatment [95].

Subsequently, *L. major*-infected patients were treated with an Amphocil<sup>®</sup> dispersion in 5% ethanol in a prospective placebo-controlled study. A significant improvement was demonstrated in Amphocil<sup>®</sup>-treated lesions compared with the placebo-treated lesions. This formulation promoted complete healing of all lesions with no evidence of relapse on follow-up visits [96]. This modality was also used for the topical treatment of an infant patient who had not responded to the topical application of a PA ointment [97], resulting in resolution of the skin lesions and absence of local or systemic side effects.

Considering that the development of lipid nanocarriers loaded with AmB for the topical treatment of CL still remains an opened question, a cationic nanoemulsion was developed and its potential for drug topical delivery was investigated through studies with dermal membranes, a damaged skin model [98]. Cationic nanocarriers were selected as carriers of drugs, since these systems have been used for improved skin delivery [90]. Other lipid nanocarriers (Amphocil<sup>®</sup> in water or in ethanol, Fungizone<sup>®</sup>, a Fungizone<sup>®</sup>-Lipofundin<sup>®</sup> mixture – FLmix) were also evaluated. Regardless of the formulation, the AmB permeation across the dermal membrane was low. In contrast, AmB penetration into the dermal membrane from Amphocil<sup>®</sup> (aqueous or hydroalcoholic) was significantly higher than that observed for other formulations, while the dermal uptake either from FLmix or nanoemulsion was lower

(Table 4). Amphocil<sup>®</sup> provided the best results, highlighted by its high improvement of dermal penetration of AmB [98].

## 6. Conclusion

Topical treatment is an attractive alternative for CL, offering significant advantages over systemic therapy. As detailed in this review, the most important drugs utilized in the topical treatment of CL include PA, imiquimod, AmB, miltefosine, and buparvaquone. Among these, PA is the most commonly studied, followed by AmB, which is useful as the second-line therapy for systemic treatment of CL. Animal and clinical studies using conventional topical formulations containing PA showed variable results, which can be attributed to low skin penetration of PA, a highly hydrophilic drug. Although PA penetration across highly damaged skin has been enhanced from hydrophilic formulations, PA permeation across the intact or partially damaged skin was still quite low. Liposomes loaded with PA improved drug skin penetration in these conditions. In agreement, PA encapsulated in liposomes has shown enhanced efficacy for the treatment of experimental CL. Topical use of AmB is also quite limited because of its low skin penetration. In this context, lipid nanocarriers can also improve topical treatment since they can enhance drug skin penetration. Among these, Amphocil<sup>®</sup> has provided the best results, highlighted by its high improvement in dermal penetration and antileishmanial activity of AmB. In synthesis, the evidence gathered from tests performed with topical formulations containing PA or AMB, two rather distinct chemical entities, indicate that lipid nanocarriers may be useful to circumvent drug properties and skin permeability, the two main aspects that limit the efficacy of CL topical treatment.

## 7. Expert opinion

The topical pharmacotherapy of CL includes a variety of drugs with well-established antileishmanial activity; however, an effective topical treatment of CL using these antileishmanial agents still remains a great challenge. PA is the most commonly studied drug for the topical treatment of CL, followed by Imiquimod, AmB, and buparvaquone. Miltefosine has been used mainly for oral administration, and concerns about drug resistance, which could be even greater after topical application, constitute a major limitation for its use in the topical therapy of CL. Imiquimod occupies a prominent place in combination with antileishmanial drugs, such as pentavalent antimony, aiming mainly at shortening the antimony therapy course. However, its efficacy when used alone is usually low.

As previously pointed out, PA is the drug most frequently studied in the topical treatment of CL, although its physicochemical properties (high hydrophilicity and high molecular weight) are not considered ideal for skin penetration. One possible explanation for the promising findings obtained with topical PA could be the high penetration of this

hydrophilic drug into and across damaged skin (open lesions), in which the barrier properties of the epidermis have been completely lost. Moreover, some disappointing results have been attributed to low skin delivery of the drug and the use of MBCL has been proposed as an alternative to improve PA skin penetration, although this has not been experimentally investigated.

The majority of studies evaluating the efficacy of PA for the topical treatment of CL used conventional drug delivery systems such as ointments, creams, and hydrophilic gels. Few studies have compared the efficacy among these systems, although the PA delivery across the damaged skin has been improved with the use of hydrophilic formulations (creams and gels). However, PA penetration into and across intact or partially damaged (without SC) pig skin was still very low, regardless of the formulation tested and this could partially explain some disappointing results observed in clinical trials. It has not been investigated whether conventional skin penetration enhancers (ethanol, fatty acids, etc.) may enhance drug penetration and, consequently, the antileishmanial activity of topical PA, has not yet been investigated. Nanocarriers have demonstrated the capacity to increase drug skin penetration, when compared with conventional formulations, and, therefore, could also provide a credible alternative for the topical treatment of the CL. Among the nanocarriers for the topical treatment of CL, liposomes are considered the most promising and several studies have reported the advantages of these systems when compared with conventional formulations.

Liposomes are known to enhance skin penetration and pharmacological activity of highly hydrophilic drugs with a high molecular weight. In agreement, liposomes loaded with PA increased drug penetration into and across intact and partially damaged skin when compared with conventional formulations. The very high water solubility of PA makes liposomes the most suitable colloidal carrier for achieving maximal drug-to-carrier ratio. Moreover, leakage of the encapsulated drug from the internal to the external aqueous phase is expected to be low due to the high hydrophilicity of PA. Although preliminary studies of efficacy in experimentally infected mice have shown the potential of these systems, this alternative should be investigated further in different models. Moreover, progress towards clinical trials is still lacking. As previously pointed out, PA-loaded topical liposomes combined with other antileishmanial drugs, administered orally or parenterally, may also represent an effective alternative to be investigated for an effective treatment for CL.

Costs and technological problems related to production of liposomes must be considered in the rational design of these formulations. One limitation is the fact that their fabrication often requires the use of organic solvents; thus, further investigation is needed to allow their large-scale production at lower costs. This is a serious limitation for the treatment of such a neglected disease as CL, which depends essentially on

therapeutic options at low costs. Alternative procedures have been described for the production of liposomes without using organic solvents as well as ways in which this could be applied to PA (Jaari and coworkers).

Although AmB is a second-line therapy useful for CL treatment, few studies have described the topical use of this antileishmanial drug. This can be attributed to the low skin penetration of such a highly hydrophobic drug with a high molecular weight. Lipid nanocarriers are often used for parenteral administration of AmB and these systems have also been described for the topical treatment of CL. Among these systems, Amphocil<sup>®</sup> occupies a prominent place in that it improved the skin penetration and antileishmanial activity of AmB when compared with other nanocarriers. Curiously, the liposomal form of AmB (Ambisome<sup>®</sup>) has not been tested for the topical treatment of CL and this should be regarded as an interesting issue in future investigations.

CL may not be taken as a single disease entity, as it is caused by rather distinct *Leishmania* species worldwide. Therefore, several factors, including the considerable variability in drug sensitivity among the infecting *Leishmania* species, ability to disseminate to distant body sites, and problems related to skin delivery, may affect treatment effectiveness. These aspects should be taken into consideration when interpreting efficacy results of chemotherapeutic studies. The variable sensitivity among species can be overcome, at least in part, by combined therapy. The usefulness of this approach, as well as that of the combination of a PA topical gel and oral miltefosine, was demonstrated experimentally for the treatment of CL. Nevertheless, the effectiveness of this approach and that of the combination between PA and other topical antileishmanial drugs in other experimental models awaits further investigation. Finally, it is highlighted that the conceptual scenario for the treatment of CL has changed significantly in the past few decades with the availability of additional drugs, routes of administration, and the knowledge of species diversity and host immune responses. This has broadened the alternatives for investigations, and in this context the results obtained with combined therapy and alternative delivery systems offer promising perspectives on improving the topical treatment of CL.

## Acknowledgement

The authors thank Glenn Hawes from the American Language Program of the University of Georgia for editing this manuscript.

## Declaration of interest

This work was supported by the "Minas Gerais State Agency for Research and Development" (FAPEMIG) and Brazilian agencies CAPES and CNPq. AP Fernandes is recipient of the CNPq research fellowship.

## Bibliography

Papers of special note have been highlighted as either of interest (●) or of considerable interest (●●) to readers

1. Sacks DL. Metacyclogenesis in *Leishmania* promastigotes. *Exp Parasitol* 1989;69:100-3
2. Kaye P, Scott P. Leishmaniasis: complexity at the host-pathogen interface. *Nat Rev Microbiol* 2011;9:604-15
3. Schriefer A, Wilson ME, Carvalho EM. Recent developments leading toward a paradigm switch in the diagnostic and therapeutic approach to human leishmaniasis. *Curr Opin Infect Dis* 2008;21:483-8
4. Grimaldi G Jr, Tesh RB, McMahon-Pratt D. A review of the geographic distribution and epidemiology of leishmaniasis in the New World. *Am J Trop Med Hyg* 1989;41:687-725
5. Berman J. Visceral leishmaniasis in the New World & Africa. *Indian J Med Res* 2006;123:289-94
6. Garnier T, Croft SL. Topical treatment for cutaneous leishmaniasis. *Curr Opin Investig Drugs* 2002;3:538-44
- **An excellent review of the key drugs used in the topical treatment of cutaneous leishmaniasis, including its chemical properties and some rational approaches to enhance skin absorption.**
7. Berman J. Clinical status of agents being developed for leishmaniasis. *Expert Opin Investig Drugs* 2005;14:1337-46
- **An excellent review of the drugs used in the treatment of visceral and cutaneous leishmaniasis, including those still in clinical development.**
8. Soto JM, Toledo JT, Gutierrez P, et al. Treatment of cutaneous leishmaniasis with a topical antileishmanial drug (WR279396): phase 2 pilot study. *Am J Trop Med Hyg* 2002;66:147-51
9. Berman J. Current treatment approaches to leishmaniasis. *Curr Opin Infect Dis* 2003;16:397-401
10. Armijos RX, Weigel MM, Calpovina M, et al. Comparison of the effectiveness of two topical paromomycin treatment versus meglumine antimoniate for new world cutaneous leishmaniasis. *Acta Trop* 2004;91:153-60
11. Santos AM, Noronha EF, Ferreira LA, et al. Efeito de uma formulacao hidrofílica de paromomicina topica na leishmaniose cutanea em pacientes com contra-indicacoes de tratamento com antimonial pentavalente. *Rev Soc Bras Med Trop* 2008;41:444-8
12. Ben-Salah A, Buffet PA, Morizot G, et al. WR279,396, a third generation aminoglycoside ointment for the treatment of *Leishmania* major cutaneous leishmaniasis: a phase 2, randomized, double Blind, placebo controlled study. *Plos Negl Trop Dis* 2009;3:432
13. Neal RA, Murphy AG, Oliaro P, Croft SL. Aminosidine ointments for the treatment of experimental cutaneous. *Trans R Soc Trop Med Hyg* 1994;88:223-35
14. Asilian A, Jalayer T, Whitworth JAG, et al. A randomized placebo-controlled trial of a two-week regimen of aminosidine (paromomycin) ointment for treatment of cutaneous leishmaniasis in Iran. *Am J Trop Med Hyg* 1995;53:648-51
15. Barry BW. Novel mechanisms and devices to enable successful transdermal drug delivery. *Eur J Pharm Sci* 2001;14:101-14
16. Papakostas D, Rancan F, Sterry W, et al. Nanoparticles in dermatology. *Arch Dermatol Res* 2011;303:533-50
17. Pierre MBR, Costa ISM. Liposomal systems as drug delivery vehicles for dermal and transdermal applications. *Arch Dermatol Res* 2011;303:607-21
18. El Maghraby GM, Barry BW, William SD. Liposomes and skin: from drug delivery to model membranes. *Eur J Pharm Sci* 2008;34:203-22
19. Carneiro G, Santos DCM, Oliveira MC, et al. Topical delivery and in vivo antileishmanial activity of paromomycin-loaded liposomes for treatment of cutaneous leishmaniasis. *J Liposome Res* 2010;20:16-23
- **This article shows the improved skin penetration and antileishmanial activity in experimental model of PA-loaded liposomes.**
20. Chiang B, Essick E, Ehringer W, et al. Enhancing skin wound healing by direct delivery of intracellular adenosine triphosphate. *Am J Surg* 2007;193:213-18
21. Lee JP, Jalili RB, Tredget EE, et al. Antifibrogenic effects of liposome-encapsulated IFN-2b cream on skin wounds in a fibrotic rabbit ear model. *J Interf Cytok Res* 2005;25:627-31
22. Langer S, Botteck NM, Bosse B, et al. Effect of polyvinylpyrrolidone-iodine liposomal hydrogel on wound microcirculation in SKH1-hr hairless mice. *Eur Surg Res* 2006;38:27-34
23. Sundar S, Chakravarty J. Paromomycin in the treatment of leishmaniasis. *Expert Opin Investig Drugs* 2008;17:787-94
24. Fernandez MM, Malchiode EL, Algranati ID. Differential effects of paromomycin on ribosomes of *Leishmania mexicana* and mammalian cells. *Antimicrob Agents Chemother* 2011;55:86-93
25. Kanyok TP, Killian AD, Rodvold KA, Danziger LH. Pharmacokinetics of intramuscularly administered aminosidine in healthy subjects. *Antimicrob Agents Chemother* 1997;41:982-6
26. Sundar S, Jha TK, Thakur CP, et al. Injectable paromomycin for visceral leishmaniasis in India. *N Engl J Med* 2007;356:2571-81
27. Aguiar MG, Silva D, Nunan FA, et al. Combined topical paromomycin and oral miltefosine treatment of mice experimentally infected with *Leishmania* (Leishmania) major leads to reduction in both lesion size and systemic parasite burdens. *J Antimicrob Chemother* 2009;64:1234-40
- **Investigation on the activity of the combination of topical PA gel and oral miltefosine for the treatment of experimental CL.**
28. Paris C, Louseau P, Bories C, Breard J. Miltefosine induces apoptosis-like death in *Leishmania donovani* promastigotes. *Antimicrob Agents Chemother* 2004;48:852-9
29. Croft SL, Engel J. Miltefosine – discovery of the antileishmanial activity of phospholipid derivatives. *Trans R Soc of Trop Med Hyg* 2006;100(Suppl 1):1-4

30. Rakotomanga M, Blanc S, Gaudin K, et al. Miltefosine affects lipid metabolism in *Leishmania donovani* promastigotes. *Antimicrob Agents Chemother* 2007;51:1425-30
31. Wadhone P, Maiti M, Agarwal R, et al. Miltefosine promotes IFN-gamma-dominated antileishmanial immune response. *J Immunol* 2009;182:7146-54
32. Soto J, Toledo J, Gutierrez P, et al. Treatment of american cutaneous leishmaniasis with miltefosine, an oral agent. *Clin Infect Dis* 2001;33:E57-61
33. Kaminsky R. Miltefosine zentaris. *Current Opin Investig Drugs* 2002;3:550-4
34. Berman J. Treatment of leishmaniasis with miltefosine: 2008 status. *Expert Opin Drug and Metab Toxicol* 2008;4:1209-12
35. Golenser J, Frakenburg S, Ehrenfreund T, Domb AJ. Efficacious treatment of experimental leishmaniasis with amphotericin B-Arabinogalactan water-soluble derivatives. *Antimicrob Agents Chemother* 1999;43:2209-14
36. Baginski M, Czuj B. Amphotericin B and Its New Derivatives – Mode of Action. *Curr Drug Metab* 2009;10:459-69
37. Croft S, Olliaro P. Leishmaniasis chemotherapy: challenges and opportunities. *Clin Microbiol Infect* 2011;17:1478-83
- **An excellent recent review on the drugs used in the treatment of leishmaniasis.**
38. Cohen BE. Amphotericin B toxicity and lethality: a tale of two channels. *Int J Pharm* 1998;162:95-106
39. Vyas SP, Gupta S. Optimizing efficacy of amphotericin B through nanomodification. *Int J Nanomedicine* 2006;1:417-32
40. Erbagci Z, Tuncel AA, Erkilic S, Zer Y. Successful treatment of antifungal- and cryotherapy-resistant subcutaneous hyalohyphomycosis in an immunocompetent case with topical 5% imiquimod cream. *Mycopathologia* 2005;159:521-6
41. Arevalo I, Ward B, Miller R, et al. Successful treatment of drug-resistant cutaneous Leishmaniasis in humans by use of imiquimod, an immunomodulator. *Clin Infect Dis* 2001;33:1847-51
42. El-On J, Bazarsky E, Sneir R. *Leishmania major*: in vitro and in vivo anti-leishmanial activity of paromomycin ointment (Leshcutan) combined with the immunomodulator imiquimod. *Exp Parasitol* 2007;116:156-62
43. Slade HB, Owens ML, Tomai MA, Miller RJ. Imiquimod 5% cream (Aldara™). *Exp Opin Invest Drugs* 1998;7:437-49
44. Sauder DN. Imiquimod: modes of action. *Br J Dermatol* 2003;149:5-8
45. Ganier T, Mantyla A, Jarvinen T, et al. In vivo studies on the antileishmanial activity of buparvaquone and its prodrugs. *J Antimicrob Chemother* 2007;60:802-10
46. Seifert K. Structures, targets and recent approaches in antileishmanial drug discovery and development. *Open Med Chem J* 2011;5:31-9
47. Moser K, Kriwet K, Naik A, et al. Passive skin penetration enhancement and its quantification in vitro. *Eur J Pharm Biopharm* 2001;52:103-12
- **An excellent review on the mechanisms and alternatives for skin penetration enhancement.**
48. Proksch E, Brandner JM, Jensen JM. The skin: an indispensable barrier. *Exp Dermatol* 2008;17:1063-72
49. Fang JY, Leu YL. Prodrug strategy for enhancing drug delivery via skin. *Curr Drug Discov Technol* 2006;3:211-24
50. Trommer H, Neubert RH. Overcoming the stratum corneum: the modulation of skin penetration. A review. *Skin Pharmacol Physiol* 2006;19:106-21
51. Reithinger R, Dujardin JC, Louzir H, et al. Cutaneous leishmaniasis. *Lancet Infect Dis* 2007;7:581-96
52. Neuber H. Leishmaniasis. *J Dtsch Dermatol Ges* 2008;6:754-652
53. Gattu S, Maibach HI. Enhanced absorption through damaged skin: an overview of the in vitro human model. *skin. Pharmacol Physiol* 2010;23:171-6
54. Markle WH, Makhoul K. Cutaneous Leishmaniasis: recognition and Treatment. *Am Fam Physician* 2004;69:1455-60
55. El-On J, Jacobs GP, Witztrum E, Greenblatt CL. Development of topical treatment for cutaneous leishmaniasis caused by *Leishmania major* in experimental animals. *Antimicrob Agents Chemother* 1984;26:745-51
- **A historic paper: the first report of a successful topical treatment with an ointment containing PA in experimentally infected mice.**
56. El-On J, Jacobs GP, Weinrauch L. Topical chemotherapy of cutaneous leishmaniasis. *Parasitol Today* 1988;4:76-81
57. Carter KC, Alexander J, Baille AJ. Studies on the topical treatment of experimental cutaneous leishmaniasis: the therapeutic effect of methyl benzethonium chloride and the aminoglycosides, gentamicin and paromomycin. *Ann Trop Med Parasitol* 1989;88:233-9
58. Grogl M, Schuster BG, Ellis WY, Berman JD. Successful topical treatment of murine cutaneous leishmaniasis with a combination of paromomycin (aminosidine) and gentamicin. *J Parasitol* 1999;85:354-9
59. El-On J, Livshin R, Even-Paz Z, et al. Topical treatment of cutaneous leishmaniasis. *J Invest Dermatol* 1986;87:1110-86
- **The first report on the treatment of CL in patients using an ointment containing PA and MBCL. Patients were treated topically for 10 days and 94% of the treated lesions healed with little or no scarring.**
60. Soto J, Hernandez N, Mejia H, et al. Successful treatment of new world cutaneous leishmaniasis with a combination of topical paromomycin/ methylbenzethonium chloride and injectable meglumine antimoniate. *Clin Infect Dis* 1995;20:47-51
61. Soto J, Fuya P, Herrera R, Berman J. Topical paromomycin/ methylbenzethonium chloride plus parenteral meglumine antimoniate as treatment for american cutaneous leishmaniasis: controlled study. *Clin Infect Dis* 1998;26:56-8

62. Arana BA, Mendoza CE, Rizzo NR, Kroeger A. Randomized, controlled, double-blind trial of topical treatment of cutaneous leishmaniasis with paromomycin plus methylbenzethonium chloride ointment in Guatemala. *Am J Trop Med Hyg* 2001;65:466-70
63. Bryceson ADM, Murphy A, Moody AH. Treatment of "Old World" cutaneous leishmaniasis with aminosidine ointment: results of an open study in London. *Trans R Soc Trop Med Hyg* 1994;88:226-8
64. Ben-Salah A, Zakraoui H, Zaatour A, et al. A randomized placebo-controlled trial in Tunisia treating cutaneous leishmaniasis with paromomycin ointment. *Am J Trop Med Hyg* 1995;53:162-6
65. Asilian A, Jalayer T, Nilforooshzadeh M, et al. Treatment of cutaneous leishmaniasis with aminosidine (paromomycin) ointment: double-blind, randomized trial in the Islamic Republic of Iran. *Bull World Health Organ* 2003;81:353-9
66. Castro GA, Nascimento DS, Nunan EA, et al. In vitro skin permeation of paromomycin from topical formulations across normal and stripped hairless mouse skin. *S. T. P. Pharm Sci* 2003;13:203-8
67. Simon GA, Maibach HI. The pig as an experimental animal model of percutaneous permeation in man: qualitative and quantitative observations – an overview. *Skin Pharmacol Appl skin Physiol* 2000;13:229-34
68. Jacobi U, Kaiser M, Toll R, et al. Porcine ear skin: an in vitro model for human skin. *Skin Res Technol* 2007;13:19-24
69. Walker M, Hulme TA, Rippon MG, et al. In Vitro models for the percutaneous delivery of active tissue repair agents. *J Pharm Sci* 1997;86:1379-84
70. Grevelink AS, Lerner EA. Leishmaniasis. *J Am Acad Dermatol* 1996;34:257-72
71. Lalor CB, Flynn GL, Weiner N. Formulation factors affecting release of drug from topical formulations. II. Effect of solubility on in vitro delivery of a series of n-alkyl p-aminobenzoates. *J Pharm Sci* 1995;84:673-6
72. Goncalves GS, Fernandes AP, Souza RCC, et al. Activity of a paromomycin hydrophilic formulation for tropical treatment of infections by *Leishmania (Leshmania) amazonensis* and *Leishmania (Vianna) braziliensis*. *Acta Trop* 2005;93:161-7
73. Mussi SV, Fernandes AP, Ferreira LA. Comparative study of the efficacy of formulations containing fluconazole or paromomycin for topical treatment of infections by *Leishmania (Leishmania) major* and *Leishmania (Leishmania) amazonensis*. *Parasitol Res* 2007;100:1221-6
74. Aguiar MG, Pereira AM, Fernandes AP, Ferreira LA. Reductions in skin and systemic parasite burdens as a combined effect of topical paromomycin and oral miltefosine treatment of mice experimentally infected with *Leishmania (Leishmania) amazonensis*. *Antimicrob Agents Chemother* 2010;54:4699-704
75. Coderch L, Fonollosa J, De Pera M, et al. Influence of cholesterol on liposome fluidity by EPR. Relationship with percutaneous absorption. *J Control Release* 2000;68:85-95
76. Bouwstra JA, Honeywell-Nguyen PL. Skin structure and mode of action of vesicles. *Adv Drug Deliv Rev* 2002;54(Suppl 1):S41-55
77. Kim MK, Chung SJ, Lee MH, et al. Targeted and sustained delivery of hydrocortisone to normal stratum corneum-removed skin without enhanced skin absorption using a liposome gel. *J Control Release* 1997;46:243-51
78. Mohebbi M, Fotouhi A, Hoosmand B, et al. Comparison of miltefosine and meglumine antimoniate for the treatment of zoonotic cutaneous leishmaniasis (ZCL) by a randomized clinical trial in Iran. *Acta Trop* 2007;103:33-40
79. Soto J, Rea J, Balderrama M, et al. Efficacy of miltefosine for Bolivian cutaneous leishmaniasis. *Am J Trop Med Hyg* 2008;78:210-11
80. Schmidt-Ott R, Klenner T, Overath P, Aebischer T. Topical treatment with hexadecylphosphocholine (Miltex®) efficiently reduces parasite burden in experimental cutaneous leishmaniasis. *Trans R Soc Trop Med Hyg* 1999;93:85-90
81. Buates S, Matlashewski G. Treatment of experimental leishmaniasis with the immunomodulators Imiquimod and S-28463: efficacy and mode of action. *J Infect Dis* 1999;179:1485-94
82. Arevalo I, Tulliano G, Quispe A, et al. Role of imiquimod and parenteral meglumine antimoniate in the initial treatment of cutaneous leishmaniasis. *Clin Infect Dis* 2007;44:1549-54
83. Al-Mutairi N, Alshiltawy M, El Khalawany M, et al. Treatment of Old World cutaneous leishmaniasis with dapsone, itraconazole, cryotherapy, and imiquimod, alone and in combination. *Int J Dermatol* 2009;48:862-9
84. Miranda-Verastegui C, Tulliano G, Gyorkos TW, et al. First-Line therapy for human cutaneous leishmaniasis in Peru using the TLR7 agonist imiquimod in combination with pentavalent antimony. *PLoS Negl Trop Dis* 2009;3:e491
85. Meymandi SS, Javadi A, Dabiri S, et al. Comparative histological and immunohistochemical changes of dry type cutaneous leishmaniasis after administration of meglumine antimoniate, imiquimod or combination therapy. *Arch Iran Med* 2011;14:238-43
86. Seeberger J, Daoud S, Pammer J. Transient effect of topical treatment of cutaneous leishmaniasis with imiquimod. *Int J Dermatol* 2003;42:576-9
87. Mantyla A, Garnier T, Rautio J, et al. Synthesis, in vitro evaluation, and antileishmanial activity of water-soluble prodrugs of buparvaquone. *J Med Chem* 2004;47:188-95
88. Garnier T, Mantyla A, Jarvinen T, et al. Topical buparvaquone formulations for the treatment of cutaneous leishmaniasis. *J Pharm Pharmacol* 2007;59:41-9
89. Samad A, Sultana Y, Aqil M. Liposomal drug delivery systems: an update review. *Curr Drug Deliv* 2007;4:297-305
90. Wu X, Landfester K, Musyanovych A, Guy RH. Disposition of charged nanoparticles after their topical application to the skin. *Skin Pharmacol Physiol* 2010;23:117-23
91. Solans C, Izquierdo P, Nolla J, et al. Nano-emulsions. *Curr Opin Coll Int Science* 2005;10:102-10
92. Ferreira LS, Ramaldes GA, Nunan EA, Ferreira LA. In vitro skin permeation and retention of paromomycin from liposomes for topical treatment of the

- cutaneous leishmaniasis. *Drug Dev Ind Pharm* 2004;30:289-96
- **The first report showing enhanced skin penetration of hydrophilic PA from liposomes.**
93. Jaafari MR, Bavarsad N, Bazzaz BSF, et al. Effect of topical liposomes containing paromomycin sulfate in the course of *Leishmania major* infection in susceptible BALB/c mice. *Antimicrob Agents Chemother* 2009;53:2259-65
  - **The first report showing the effects of topical liposomes loaded with PA for the treatment of experimental CL.**
  94. Egitto EST, Araujo IB, Damasceno BPG, Price JC. Amphotericin B/emulsion admixture interactions: an approach concerning the reduction of amphotericin B toxicity. *J Pharm Sci* 2002;91:2354-66
  95. Frankenburg S, Glick D, Klaus S, Barenholz Y. Efficacious topical treatment for murine cutaneous leishmaniasis with ethanolic formulations of amphotericin B. *Antimicrob Agents Chemother* 1998;42:3092-6
  96. Vardy D, Barenholz Y, Naftoliev N, et al. Efficacious topical treatment for human cutaneous leishmaniasis with ethanolic lipid amphotericin B. *Trans R Soc Trop Med Hyg* 2001;95:184-6
  97. Zvulunov A, Cagnano E, Frankenburg S, et al. Topical treatment of persistent cutaneous leishmaniasis with ethanolic lipid amphotericin B. *Pediatr Infect Dis J* 2003;22:567-9
  98. Santos CM, Oliveira RB, Arantes VT, et al. Amphotericin B-loaded Nanocarriers for Topical Treatment of Cutaneous Leishmaniasis: Development, Characterization, and In Vitro Skin Permeation Studies. *J Biomed Nanotechnol* 2012;8:322-9. Copyright © American Scientific Publishers.
  - **This paper describes the preparation and characterization of AmB-loaded nanocarriers. It also investigates the topical delivery of AMB from the nanosystems by conducting permeation studies.**
  99. The United Pharmacopeia; the national formulary - USP 31. United States Pharmacopeial convention, Rockville; 2008
  100. British Pharmacopeia. Her Majesty's Stationary Office, London; 2005

### Affiliation

Guilherme Carneiro<sup>1</sup> MD,  
Marta Gontijo Aguiar<sup>2</sup> PhD,  
Ana Paula Fernandes<sup>3</sup> PhD &  
Lucas Antônio Miranda Ferreira<sup>†4</sup> PhD  
<sup>†</sup>Author for correspondence  
<sup>1</sup>Federal University of Minas Gerais,  
Faculty of Pharmacy,  
Belo Horizonte,  
Minas Gerais, Brazil  
<sup>2</sup>Newton Paiva University Center, Belo  
Horizonte, Minas Gerais, Brazil  
<sup>3</sup>Federal University of Minas Gerais, Faculty of  
Pharmacy, Belo Horizonte, Minas Gerais, Brazil  
<sup>4</sup>Federal University of Minas Gerais, Department  
of Pharmaceutics,  
Faculty of Pharmacy, Av Antônio Carlos,  
6627, Campus Pampulha. CEP 31270-901,  
Belo Horizonte, Minas Gerais, Brazil  
Tel: +55 31 3409 6939; Fax: +55 31 3409 6830;  
E-mail: lucas@farmacia.ufmg.br